Package: abn (via r-universe)

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Title Modelling Multivariate Data with Additive Bayesian Networks

Version 3.1.2

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Description The 'abn' R package facilitates Bayesian network analysis, a probabilistic graphical model that derives from empirical data a directed acyclic graph (DAG). This DAG describes the dependency structure between random variables. The R package 'abn' provides routines to help determine optimal Bayesian network models for a given data set. These models are used to identify statistical dependencies in messy, complex data. Their additive formulation is equivalent to multivariate generalised linear modelling, including mixed models with independent and identically distributed (iid) random effects. The core functionality of the 'abn' package revolves around model selection, also known as structure discovery. It supports both exact and heuristic structure learning algorithms and does not restrict the data distribution of parent-child combinations, providing flexibility in model creation and analysis. The 'abn' package uses Laplace approximations for metric estimation and includes wrappers to the 'INLA' package. It also employs 'JAGS' for data simulation purposes. For more resources and information, visit the 'abn' website.

License GPL $(>= 3)$

URL <https://r-bayesian-networks.org/>,

<https://github.com/furrer-lab/abn>

BugReports <https://github.com/furrer-lab/abn/issues>

Depends R $(>= 4.0.0)$

- Imports doParallel, foreach, graph, lme4, mclogit, methods, nnet, Rcpp, Rgraphviz, rjags, stringi,
- **Suggests** bookdown, boot, brglm, devtools $(>= 2.4.5)$, ggplot2, gridExtra, INLA, knitr, Matrix, MatrixModels (>= 0.5.3), microbenchmark, R.rsp, RhpcBLASctl, rmarkdown, testthat (>= 3.0.0), entropy, moments, R6

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LinkingTo Rcpp, RcppArmadillo

VignetteBuilder knitr

Additional_repositories <https://inla.r-inla-download.org/R/stable/>

Config/testthat/edition 3

Encoding UTF-8

LazyData TRUE

Roxygen list(markdown = TRUE)

RoxygenNote 7.3.1

SystemRequirements pkg-config, cmake, gsl, jpeg, gdal, geos, proj, udunits-2, openssl, libcurl, jags

Repository https://furrer-lab.r-universe.dev

RemoteUrl https://github.com/furrer-lab/abn

RemoteRef HEAD

RemoteSha 3ff721d6539712714743950c14a220dd44284878

Contents

AIC.abnFit *Print AIC of objects of class* abnFit

Description

Print AIC of objects of class abnFit

Usage

```
## S3 method for class 'abnFit'
AIC(object, digits = 3L, verbose = TRUE, ...)
```
Arguments

Value

prints the AIC of the fitted model.

Print BIC of objects of class abnFit

Usage

```
## S3 method for class 'abnFit'
BIC(object, digits = 3L, verbose = TRUE, ...)
```
Arguments

Value

prints the BIC of the fitted model.

Description

Allow the user to set restrictions in the [buildScoreCache](#page-0-0) for both the Bayesian and the MLE approach. Control function similar to [fit.control](#page-18-1).

Usage

```
build.control(
  method = "bayes",
  max.mode.error = 10,
  mean = 0,
  prec = 0.001,loggam.shape = 1,loggam.inv.scale = <math>5e-05</math>,max.iters = 100,epsabs = 1e-07,
  error.verbose = FALSE,
  trace = \mathfrak{D}L,
  epsabs.inner = 1e-06,
```
build.control 5

```
max.iters.inner = 100,
finite.step.size = 1e-07,
hessian.params = c(1e-04, 0.01),
max.iters.hessian = 10,
max. hessian.crror = 0.5,factor.brent = 100,
maxiters.hessian.brent = 100,
num.intervals.brent = 100,
n.grid = 250,
ncores = 1,
cluster.type = "FORK",
max.irls = 100,tol = 1e-08,tolPwrs = 1e-07,
check.rankX = "message+drop.cols",
check.scaleX = "message+rescale",
check.conv.grad = "message",
check.conv.singular = "message",
check.conv.hess = "message",
xtol_abs = 1e-06,
ftol_abs = 1e-06,
trace.mblogit = FALSE,
catcov.mblogit = "free",
epsilon = 1e-06,
seed = 9062019L
```
Arguments

 \mathcal{L}

Details

Parallelization over all children is possible via the function foreach of the package doParallel. ncores=0 or ncores=1 use single threaded foreach. ncores=-1 uses all available cores but one.

Value

Named list according the provided arguments.

See Also

[fit.control](#page-18-1).

Other buildScoreCache: [buildScoreCache\(](#page-0-0))

Examples

```
ctrlmle <- abn::build.control(method = "mle",
                        ncores = 0,
                        cluster.type = "PSOCK",
                        max.irls = 100,tol = 10^{\circ} - 11,
                        tolPwrss = 1e-7,
                        check.rankX = "message+drop.cols",
                        check.scaleX = "message+rescale",
                        check.conv.grad = "message",
                        check.conv.singular = "message",
                        check.conv.hess = "message",
                        xtol_abs = 1e-6,
                        ftol_abs = 1e-6,
                        trace.mblogit = FALSE,
                        catcov.mblogit = "free",
                        epsilon = 1e-6,
                        seed = 9062019L)
ctrlbayes <- abn::build.control(method = "bayes",
                           max.mode.error = 10,
                           mean = 0, prec = 0.001,
                           loggam.shape = 1,loggam.inv.scale = 5e-05,
                           max.iters = 100,
                           epsabs = 1e-07,
                           error.verbose = FALSE,
                           epsabs.inner = 1e-06,
                           max.iters.inner = 100,
                           finite.step.size = 1e-07,
                           hessian.params = c(1e-04, 0.01),
                           max.iters.hessian = 10,
```

```
max. hessian. error = 0.5,factor.brent = 100,
maxiters.hessian.brent = 100,
num.intervals.brent = 100,
tol = 10^{\circ} - 8,
seed = 9062019L)
```
check.valid.fitControls

Simple check on the control parameters

Description

Simple check on the control parameters

Usage

```
check.valid.fitControls(control, method = "bayes", verbose = FALSE)
```
Arguments

Value

list with all control arguments with respect to the method but with new values.

coef.abnFit *Print coefficients of objects of class* abnFit

Description

Print coefficients of objects of class abnFit

Usage

```
## S3 method for class 'abnFit'
coef(object, digits = 3L, verbose = TRUE, ...)
```
Arguments

Value

prints the coefficients of the fitted model.

compareDag *Compare two DAGs or EGs*

Description

Function that returns multiple graph metrics to compare two DAGs or essential graphs, known as confusion matrix or error matrix.

Usage

compareDag(ref, test, node.names = NULL, checkDAG = TRUE)

Arguments

Details

This R function returns standard Directed Acyclic Graph comparison metrics. In statistical classification, those metrics are known as a confusion matrix or error matrix.

Those metrics allows visualization of the difference between different DAGs. In the case where comparing TRUTH to learned structure or two learned structures, those metrics allow the user to estimate the performance of the learning algorithm. In order to compute the metrics, a contingency table is computed of a pondered difference of the adjacency matrices od the two graphs.

The ref or test can be provided using a formula statement (similar to GLM input). A typical formula is ~ node1|parent1:parent2 + node2:node3|parent3. The formula statement have to start with ~. In this example, node1 has two parents (parent1 and parent2). node2 and node3 have the same parent3. The parents names have to exactly match those given in node.names. : is the

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separtor between either children or parents, \vert separates children (left side) and parents (right side), + separates terms, . replaces all the variables in node.names.

To test for essential graphs (or graphs) in general, the test for DAG need to be switched off checkDAG=FALSE. The function compareEG() is a wrapper to compareDag(, checkDAG=FALSE).

Value

- TP True Positive
- TN True Negative
- FP False Positive
- FN False Negative
- CP Condition Positive (ref)
- CN Condition Negative (ref)
- PCP Predicted Condition Positive (test)
- PCN Predicted Condition Negative (test)

True Positive Rate

False Positive Rate

$$
= \frac{\sum TP}{\sum CP}
$$

$$
= \frac{\sum FP}{\sum CN}
$$

Accuracy

$$
= \frac{\sum TP + \sum TN}{Total population}
$$

G-measure

$$
\sqrt{\frac{TP}{TP+FP}\cdot\frac{TP}{TP+FN}}
$$

F1-Score

$$
\frac{2\sum TP}{2\sum TP + \sum FN + \sum FP}
$$

 $\sum TP$ $\sum PCF$

 $\sum FN$

Positive Predictive Value

False Ommision Rate

$$
\overline{\sum PCN}
$$

Hamming-Distance Number of changes needed to match the matrices.

References

Sammut, Claude, and Geoffrey I. Webb. (2017). Encyclopedia of machine learning and data mining. Springer.

Examples

```
test.m \leq matrix(data = c(0,1,0,0,0,0,
                           1, 0, 0, nrow = 3, ncol = 3)
ref.m \leq matrix(data = c(0,0,0,0)1,0,0,
                          1, 0, 0, nrow = 3, ncol = 3)
```
colnames(test.m) <- rownames(test.m) <- colnames(ref.m) <- colnames(ref.m) <- c("a", "b", "c")

```
unlist(compareDag(ref = ref.m, test = test.m))
```


Compare two DAGs or EGs

Description

Function that returns multiple graph metrics to compare two DAGs or essential graphs, known as confusion matrix or error matrix.

Usage

compareEG(ref, test)

Arguments

Details

This R function returns standard Directed Acyclic Graph comparison metrics. In statistical classification, those metrics are known as a confusion matrix or error matrix.

Those metrics allows visualization of the difference between different DAGs. In the case where comparing TRUTH to learned structure or two learned structures, those metrics allow the user to estimate the performance of the learning algorithm. In order to compute the metrics, a contingency table is computed of a pondered difference of the adjacency matrices od the two graphs.

The ref or test can be provided using a formula statement (similar to GLM input). A typical formula is \sim node1|parent1:parent2 + node2:node3|parent3. The formula statement have to start with \sim . In this example, node1 has two parents (parent1 and parent2). node2 and node3 have the same parent3. The parents names have to exactly match those given in node.names. : is the separtor between either children or parents, \vert separates children (left side) and parents (right side), + separates terms, . replaces all the variables in node.names.

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To test for essential graphs (or graphs) in general, the test for DAG need to be switched off checkDAG=FALSE. The function compareEG() is a wrapper to compareDag(, checkDAG=FALSE).

Value

- TP True Positive
- TN True Negative
- FP False Positive
- FN False Negative
- CP Condition Positive (ref)
- CN Condition Negative (ref)
- PCP Predicted Condition Positive (test)
- PCN Predicted Condition Negative (test)

True Positive Rate

$$
= \frac{\sum TP}{\sum CP}
$$

False Positive Rate

$$
= \frac{\sum FP}{\sum CN}
$$

Accuracy

$$
= \frac{\sum TP + \sum TN}{Total population}
$$

G-measure

$$
\sqrt{\frac{TP}{TP+FP}\cdot\frac{TP}{TP+FN}}
$$

F1-Score

$$
\frac{2\sum TP}{2\sum TP + \sum FN + \sum FP}
$$

Positive Predictive Value

$$
\frac{\sum TP}{\sum PCP}
$$

False Ommision Rate

$$
\frac{\sum FN}{\sum PCN}
$$

Hamming-Distance Number of changes needed to match the matrices.

References

Sammut, Claude, and Geoffrey I. Webb. (2017). Encyclopedia of machine learning and data mining. Springer.

Examples

```
test.m <- matrix(data = c(0,1,0,0,0,0,
                          1, 0, 0, nrow = 3, ncol = 3)
ref.m \leq matrix(data = c(0,0,0,0)1,0,0,
                         1,0,0), nrow = 3, ncol = 3)
colnames(test.m) <- rownames(test.m) <- colnames(ref.m) <- colnames(ref.m) <- c("a", "b", "c")
unlist(compareDag(ref = ref.m, test = test.m))
```
discretization *Discretization of a Possibly Continuous Data Frame of Random Variables based on their distribution*

Description

This function discretizes a data frame of possibly continuous random variables through rules for discretization. The discretization algorithms are unsupervised and univariate. See details for the complete list of discretization rules (the number of state of each random variable could also be provided).

Usage

```
discretization(data.df = NULL,
                      data.dists = NULL,
                      discretization.method = "sturges",
                      nb.states = FALSE)
```
Arguments

Details

fd Freedman Diaconis rule. IQR() stands for interquartile range. The number of bins is given by

$$
\frac{range(x) * n^{1/3}}{2 * IQR(x)}
$$

The Freedman Diaconis rule is known to be less sensitive than the Scott's rule to outlier. doane Doane's rule. The number of bins is given by

$$
1 + \log_2 n + \log_2 1 + \frac{|g|}{\sigma_g}
$$

This is a modification of Sturges' formula, which attempts to improve its performance with nonnormal data.

sqrt The number of bins is given by:

$$
\sqrt(n)
$$

cencov Cencov's rule. The number of bins is given by:

$$
n^{1/3}
$$

rice Rice' rule. The number of bins is given by:

$$
2n^{1/3}
$$

terrell-scott Terrell-Scott's rule. The number of bins is given by:

 $(2n)^{1/3}$

It is known that Cencov, Rice, and Terrell-Scott rules over-estimates k, compared to other rules due to its simplicity.

sturges Sturges's rule. The number of bins is given by:

 $1 + \log_2(n)$

scott Scott's rule. The number of bins is given by:

 $range(x)/\sigma(x)n^{-1/3}$

Value

The discretized data frame or a list containing the table of counts for each bin the discretized data frame.

table of counts for each bin of the discretized data frame.

References

Garcia, S., et al. (2013). A survey of discretization techniques: Taxonomy and empirical analysis in supervised learning. *IEEE Transactions on Knowledge and Data Engineering*, 25.4, 734-750.

Cebeci, Z. and Yildiz, F. (2017). Unsupervised Discretization of Continuous Variables in a Chicken Egg Quality Traits Dataset. *Turkish Journal of Agriculture-Food Science and Technology*, 5.4, 315- 320.

Examples

```
## Generate random variable
rv \le -rnorm(n = 100, mean = 5, sd = 2)dist <- list("gaussian")
names(dist) <- c("rv")
## Compute the entropy through discretization
entropyData(freqs.table = discretization(data.df = rv, data.dists = dist,
discretization.method = "sturges", nb.states = FALSE))
```


Description

This function empirically estimates the Shannon entropy from a table of counts using the observed frequencies.

Usage

entropyData(freqs.table)

Arguments

freqs.table a table of counts.

Details

The general concept of entropy is defined for probability distributions. The entropyData() function estimates empirical entropy from data. The probability is estimated from data using frequency tables. Then the estimates are plug-in in the definition of the entropy to return the so-called empirical entropy. A common known problem of empirical entropy is that the estimations are biased due to the sampling noise. It is also known that the bias will decrease as the sample size increases.

Value

Shannon's entropy estimate on natural logarithm scale.

integer

References

Cover, Thomas M, and Joy A Thomas. (2012). "Elements of Information Theory". John Wiley & Sons.

See Also

[discretization](#page-13-1)

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Examples

```
## Generate random variable
rv \le -rnorm(n = 100, mean = 5, sd = 2)dist <- list("gaussian")
names(dist) <- c("rv")
## Compute the entropy through discretization
entropyData(freqs.table = discretization(data.df = rv, data.dists = dist,
discretization.method = "sturges", nb.states = FALSE))
```
essentialGraph *Construct the essential graph*

Description

Constructs different versions of the essential graph from a given DAG. External function that computes essential graph of a dag Minimal PDAG: The only directed edges are those who participate in v-structure Completed PDAG: very directed edge corresponds to a compelled edge, and every undirected edge corresponds to a reversible edge

Usage

essentialGraph(dag, node.names = NULL, PDAG = "minimal")

Arguments

Details

This function returns an essential graph from a DAG, aka acyclic partially directed graph (PDAG). This can be useful if the learning procedure is defined up to a Markov class of equivalence. A minimal PDAG is defined as only directed edges are those who participate in v-structure. Whereas the completed PDAG: every directed edge corresponds to a compelled edge, and every undirected edge corresponds to a reversible edge.

The dag can be provided using a formula statement (similar to glm). A typical formula is \sim node1|parent1:parent2 + node2:node3|parent3. The formula statement have to start with \sim . In this example, node1 has two parents (parent1 and parent2). node2 and node3 have the same parent3. The parents names have to exactly match those given in node.names. : is the separator between either children or parents, | separates children (left side) and parents (right side), + separates terms, . replaces all the variables in node.names.

Value

A matrix giving the PDAG.

References

West, D. B. (2001). Introduction to Graph Theory. Vol. 2. Upper Saddle River: Prentice Hall. Chickering, D. M. (2013) A Transformational Characterization of Equivalent Bayesian Network Structures, arXiv:1302.4938.

Examples

```
dag <- matrix(c(0,0,0, 1,0,0, 1,1,0), nrow = 3, ncol = 3)
dist <- list(a="gaussian", b="gaussian", c="gaussian")
colnames(dag) <- rownames(dag) <- names(dist)
```
essentialGraph(dag)

Description

See also the C implementation ?abn::expit_cpp().

Usage

expit(x)

Arguments

x numeric with values between $[0, 1]$.

Value

numeric vector of same length as x.

expit_cpp *expit function*

Description

transform x either via the logit, or expit.

Usage

expit_cpp(x)

Arguments

x a numeric vector

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Value

a numeric vector

family.abnFit *Print family of objects of class* abnFit

Description

Print family of objects of class abnFit

Usage

S3 method for class 'abnFit' family(object, ...)

Arguments

Value

prints the distributions for each variable of the fitted model.

Description

Allow the user to set restrictions in the [fitAbn](#page-0-0) for both the Bayesian and the MLE approach. Control function similar to [build.control](#page-3-1).

Usage

```
fit.control(
  method = "bayes",
  max.mode.error = 10,
  mean = 0,
  prec = 0.001,loggam.shape = 1,loggam.inv.scale = <math>5e-05</math>,max.iters = 100,
  epsabs = 1e-07,
  error.verbose = FALSE,
  trace = \thetaL,
```

```
epsabs.inner = 1e-06,
 max.iters.inner = 100,
 finite.step.size = 1e-07,
 hessian.params = c(1e-04, 0.01),
 max.iters.hessian = 10,
 max.hessian.error = 1e-04,
 factor.brent = 100,
 maxiters.hessian.brent = 10,
 num.intervals.brent = 100,
 min.pdf = 0.001,n.grid = 250,
  std.area = TRUE,
 marginal.quantiles = c(0.025, 0.25, 0.5, 0.75, 0.975),
 max.grid.iter = 1000,marginal.node = NULL,
 marginal.param = NULL,
 variate.vec = NULL,
 ncores = 1,
 cluster.type = "FORK",
 max.irls = 100.
 tol = 1e-11,tolPwrss = 1e-07,check.rankX = "message+drop.cols",
  check.scaleX = "message+rescale",
  check.conv.grad = "message",
  check.conv.singular = "message",
  check.conv.hess = "message",
 xtol_abs = 1e-06,
  ftol_abs = 1e-06,
  trace.mblogit = FALSE,
 catcov.mblogit = "free",
 epsilon = 1e-06,
  seed = 9062019L
\lambda
```
Arguments

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loggam.inv.

...).

max.hessian.error

if the estimated log marginal likelihood when using an adaptive 5pt finite-difference rule for the Hessian differs by more than max.hessian.error from when using an adaptive 3pt rule then continue to minimize the local error by switching to the Brent-Dekker root bracketing method. Passed to .Call("fit_single_node", ...).

factor.brent if using Brent-Dekker root bracketing method then define the outer most interval end points as the best estimate of h (stepsize) from the Nelder-Mead as $h/factor.brent, h * factor.brent$). Passed to .Call("fit_single_node", ...).

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check.conv.grad character passed to [lmerControl](#page-0-0) and [glmerControl](#page-0-0) - checking the gradient of the deviance function for convergence. Defaults to message but can be one of "ignore" - skip the test; "warning" - warn if test fails; "message" - print a message if test fails; "stop" - throw an error if test fails. check.conv.singular character passed to [lmerControl](#page-0-0) and [glmerControl](#page-0-0) - checking for a singular fit, i.e. one where some parameters are on the boundary of the feasible space (for example, random effects variances equal to 0 or correlations between random effects equal to $+/- 1.0$). Defaults to message but can be one of "ignore" - skip the test; "warning" - warn if test fails; "message" - print a message if test fails; "stop" - throw an error if test fails. check.conv.hess character passed to [lmerControl](#page-0-0) and [glmerControl](#page-0-0) - checking the Hessian of the deviance function for convergence. Defaults to message but can be one of "ignore" - skip the test; "warning" - warn if test fails; "message" - print a message if test fails; "stop" - throw an error if test fails. xtol_abs Defaults to 1e-6 stop on small change of parameter value. Only for method='mle', group.var=.... Default convergence tolerance for fitted (g)lmer models is reduced to the value provided here if default values did not fit. This value here is passed to the optCtrl argument of (g) lmer (see help of [lme4::convergence\(\)](#page-0-0)). ftol_abs Defaults to 1e-6 stop on small change in deviance. Similar to xtol_abs. trace.mblogit logical indicating if output should be produced for each iteration. Directly passed to trace argument in [mclogit.control](#page-0-0). Is independent of verbose. catcov.mblogit Defaults to "free" meaning that there are no restrictions on the covariances of random effects between the logit equations. Set to "diagonal" if random effects pertinent to different categories are uncorrelated or "single" if random effect variances pertinent to all categories are identical. epsilon Defaults to 1e-8. Positive convergence tolerance ϵ that is directly passed to the control argument of mclogit::mblogit as mclogit.control. Only for method='mle', group.var=.... seed a non-negative integer which sets the seed in set. seed(seed).

Details

Parallelization over all children is possible via the function foreach of the package **doParallel**. ncores=0 or ncores=1 use single threaded foreach. ncores=-1 uses all available cores but one.

Value

a list of control parameters for the [fitAbn](#page-0-0) function.

See Also

[build.control](#page-3-1). Other fitAbn: [fitAbn\(](#page-0-0))

Examples

```
ctrlmle <- abn::fit.control(method = "mle",
                       max.irls = 100,
                       tol = 10^{\circ}-11,
                       tolPwrss = 1e-7,xtol_abs = 1e-6,
                        ftol_abs = 1e-6,
                        epsilon = 1e-6,
                       ncores = 2,
                       cluster.type = "PSOCK",
                        seed = 9062019L)
ctrlbayes <- abn::fit.control(method = "bayes",
                         mean = 0,
                         prec = 0.001,
                         loggam.shape = 1,loggam.inv.scale = <math>5e-05</math>,max.mode.error = 10,
                         max.iters = 100,epsabs = 1e-07,
                          error.verbose = FALSE,
                          epsabs.inner = 1e-06,
                         max.iters.inner = 100,
                         finite.step.size = 1e-07,
                         hessian.params = c(1e-04, 0.01),
                         max.iters.hessian = 10,
                         max.hessian.error = 1e-04,
                         factor.brent = 100,
                         maxiters.hessian.brent = 10,
                         num.intervals.brent = 100,
                         min.pdf = 0.001,n.grid = 100,
                          std.area = TRUE,
                         marginal.quantiles = c(0.025, 0.25, 0.5, 0.75, 0.975),
                         max.grid.iter = 1000,
                         marginal.node = NULL,
                         marginal.param = NULL,
                         variate.vec = NULL,
                         ncores = 1,
                         cluster.type = NULL,
                          seed = 9062019L)
```
getMSEfromModes *Extract Standard Deviations from all Gaussian Nodes*

Description

Extract Standard Deviations from all Gaussian Nodes

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Usage

getMSEfromModes(modes, dists)

Arguments

Value

named numeric vector. Names correspond to node name. Value to standard deviations.

infoDag *Compute standard information for a DAG.*

Description

This function returns standard metrics for DAG description. A list that contains the number of nodes, the number of arcs, the average Markov blanket size, the neighborhood average set size, the parent average set size and children average set size.

Usage

infoDag(object, node.names = NULL)

Arguments

node.names a vector of names if the DAG is given via formula, see details.

Details

This function returns a named list with the following entries: the number of nodes, the number of arcs, the average Markov blanket size, the neighborhood average set size, the parent average set size, and the children's average set size.

The dag can be provided using a formula statement (similar to glm). A typical formula is \sim node1|parent1:parent2 + node2:node3|parent3. The formula statement have to start with \sim . In this example, node1 has two parents (parent1 and parent2). node2 and node3 have the same parent3. The parents names have to exactly match those given in node.names. : is the separator between either children or parents, | separates children (left side) and parents (right side), + separates terms, . replaces all the variables in node.names.

Value

A named list that contains following entries: the number of nodes, the number of arcs, the average Markov blanket size, the neighborhood average set size, the parent average set size and children average set size.

References

West, D. B. (2001). Introduction to graph theory. Vol. 2. Upper Saddle River: Prentice Hall.

Examples

```
## Creating a dag:
dag <- matrix(c(\emptyset, \emptyset, \emptyset, 0, 1, \emptyset, \emptyset, 1, 1, \emptyset, 1, 0, 1, \emptyset, \emptyset), nrow = 4, ncol = 4)
dist <- list(a="gaussian", b="gaussian", c="gaussian", d="gaussian")
colnames(dag) <- rownames(dag) <- names(dist)
infoDag(dag)
plot(createAbnDag(dag = dag, data.dists = dist))
```


Description

A flexible implementation of multiple proxy for strength measures useful for visualizing the edge connections in a Bayesian Network learned from observational data.

Usage

```
linkStrength(dag,
```

```
data.df = NULL,data.dists = NULL,
method = c("mi.raw","mi.raw.pc",
           "mi.corr",
           "ls",
           "ls.pc",
           "stat.dist"),
discretization.method = "doane")
```
Arguments

a character vector giving the discretization method to use. See [discretization](#page-13-1).

Details

This function returns multiple proxies for estimating the connection strength of the edges of a possibly discretized Bayesian network's data set. The returned connection strength measures are: the Raw Mutual Information (mi.raw), the Percentage Mutual information (mi.raw.pc), the Raw Mutual Information computed via correlation (mi.corr), the link strength (1s), the percentage link strength (ls.pc) and the statistical distance (stat.dist).

The general concept of entropy is defined for probability distributions. The probability is estimated from data using frequency tables. Then the estimates are plug-in in the definition of the entropy to return the so-called empirical entropy. A standard known problem of empirical entropy is that the estimations are biased due to the sampling noise. This is also known that the bias will decrease as the sample size increases. The mutual information estimation is computed from the observed frequencies through a plug-in estimator based on entropy. For the case of an arc going from the node X to the node Y and the remaining set of parent of Y is denoted as Z.

The mutual information is defined as $I(X, Y) = H(X) + H(Y) - H(X, Y)$, where H() is the entropy.

The Percentage Mutual information is defined as $PI(X, Y) = I(X, Y)/H(Y|Z)$.

The Mutual Information computed via correlation is defined as $MI(X,Y) = -0.5 \log(1-\text{cor}(X,Y)^2)$.

The link strength is defined as $LS(X\rightarrow Y) = H(Y|Z) - H(Y|X,Z)$.

The percentage link strength is defined as $PLS(X\rightarrow Y) = LS(X\rightarrow Y) / H(Y|Z)$.

The statistical distance is defined as $SD(X,Y) = 1 - MI(X,Y) / max(H(X),H(Y)).$

Value

The function returns a named matrix with the requested metric.

References

Boerlage, B. (1992). Link strength in Bayesian networks. Diss. University of British Columbia. Ebert-Uphoff, Imme. "Tutorial on how to measure link strengths in discrete Bayesian networks." (2009).

Examples

```
# Gaussian
N < - 1000mydists <- list(a="gaussian",
                  b="gaussian",
                  c="gaussian")
a \leq -rnorm(n = N, mean = 0, sd = 1)b \le -1 + 2*rnorm(n = N, mean = 5, sd = 1)
c \le -2 + 1 \times a + 2 \times b + \text{norm}(n = N, \text{ mean } = 2, \text{ sd } = 1)mydf <- data.frame("a" = a,
                      "b" = b,
```

```
C'' = Cmycache.mle <- buildScoreCache(data.df = mydf,
                              data.dists = mydists,
                              method = "mle",max.parents = 2)
mydag.mp <- mostProbable(score.cache = mycache.mle, verbose = FALSE)
linkstr <- linkStrength(dag = mydag.mp$dag,
                       data.df = mydf,data.dists = mydists,
                       method = "ls",discretization.method = "sturges")
```
logit *Logit of proportions*

Description

See also the C implementation ?abn::logit_cpp().

Usage

logit(x)

Arguments

x numeric with values between $[0, 1]$.

Value

numeric vector of same length as x. numeric vector of same length as x.

logit_cpp *logit functions*

Description

transform x either via the logit, or expit.

Usage

logit_cpp(x)

Arguments

x a numeric vector

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Value

a numeric vector

logLik.abnFit *Print logLik of objects of class* abnFit

Description

Print logLik of objects of class abnFit

Usage

```
## S3 method for class 'abnFit'
logLik(object, digits = 3L, verbose = TRUE, ...)
```
Arguments

Value

prints the logLik of the fitted model.

mb *Compute the Markov blanket*

Description

This function computes the Markov blanket of a set of nodes given a DAG (Directed Acyclic Graph).

Usage

mb(dag, node, data.dists = NULL)

Arguments

Details

This function returns the Markov Blanket of a set of nodes given a DAG.

The dag can be provided using a formula statement (similar to glm). A typical formula is \sim node1|parent1:parent2 + node2:node3|parent3. The formula statement have to start with \sim . In this example, node1 has two parents (parent1 and parent2). node2 and node3 have the same parent3. The parents names have to exactly match those given in name. : is the separtor between either children or parents, | separates children (left side) and parents (right side), + separates terms, . replaces all the variables in name.

Value

character vector of node names from the Markov blanket.

Examples

```
## Defining distribution and dag
dist <- list(a="gaussian", b="gaussian", c="gaussian", d="gaussian",
             e="binomial", f="binomial")
dag <- matrix(c(0,1,1,0,1,0,
                0,0,1,1,0,1,
                0,0,0,0,0,0,
                0, 0, 0, 0, 0, 0,0,0,0,0,0,1,
                0,0,0,0,0,0), nrow = 6L, ncol = 6L, byrow = TRUE)
colnames(dag) <- rownames(dag) <- names(dist)
mb(dag, node = "b")mb(dag, node = c("b", "e"))mb(~a|b:c:e+b|c:d:f+e|f, node = "e", data.dists = dist)
```
miData *Empirical Estimation of the Entropy from a Table of Counts*

Description

This function empirically estimates the Mutual Information from a table of counts using the observed frequencies.

Usage

miData(freqs.table, method = c("mi.raw", "mi.raw.pc"))

Arguments

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Details

The mutual information estimation is computed from the observed frequencies through a plugin estimator based on entropy.

The plugin estimator is

$$
I(X,Y) = H(X) + H(Y) - H(X,Y)
$$

, where

 $H()$

is the entropy computed with [entropyData](#page-15-1).

Value

Mutual information estimate.

integer

References

Cover, Thomas M, and Joy A Thomas. (2012). "Elements of Information Theory". John Wiley & Sons.

See Also

[discretization](#page-13-1)

Examples

```
## Generate random variable
Y \le - rnorm(n = 100, mean = 0, sd = 2)
X < - rnorm(n = 100, mean = 5, sd = 2)
dist <- list(Y="gaussian", X="gaussian")
miData(discretization(data.df = cbind(X,Y), data.dists = dist,
                      discretization.method = "fd", nb.states = FALSE),
                      method = "mi.raw")
```
modes2coefs *Convert modes to fitAbn.mle\$coefs structure*

Description

Convert modes to fitAbn.mle\$coefs structure

Usage

modes2coefs(modes)

Arguments

modes list of modes.

Value

list of matrix arrays.

mostProbable *Find most probable DAG structure*

Description

Find most probable DAG structure using exact order based approach due to Koivisto and Sood, 2004.

Usage

```
mostProbable(score.cache, score="bic", prior.choice=1, verbose=TRUE, ...)
```
Arguments

Details

The procedure runs the exact order based structure discovery approach of Koivisto and Sood (2004) to find the most probable posterior network (DAG). The local.score is the node cache, as created using [buildScoreCache](#page-0-0) (or manually provided the same format is used). Note that the scope of this search is given by the options used in local.score, for example, by restricting the number of parents or the ban or retain constraints given there.

This routine can take a long time to complete and is highly sensitive to the number of nodes in the network. It is recommended to use this on a reduced data set to get an idea as to the computational practicality of this approach. In particular, memory usage can quickly increase to beyond what may be available. For additive models, problems comprising up to 20 nodes are feasible on most machines. Memory requirements can increase considerably after this, but then so does the run time making this less practical. It is recommended that some form of over-modeling adjustment is performed on this resulting DAG (unless dealing with vast numbers of observations), for example, using parametric bootstrapping, which is straightforward to implement in MCMC engines such as JAGS or WinBUGS. See the case studies at <https://r-bayesian-networks.org/> or the files

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provided in the package directories inst/bootstrapping_example and inst/old_vignette for details.

The parameter prior.choice determines the prior used within each node for a given choice of parent combination. In Koivisto and Sood (2004) p.554, a form of prior is used, which assumes that the prior probability for parent combinations comprising of the same number of parents are all equal. Specifically, that the prior probability for parent set G with cardinality |G| is proportional to $1/[\![n-1]\!]$ choose $|\![G]\!]$ where there are n total nodes. Note that this favors parent combinations with either very low or very high cardinality, which may not be appropriate. This prior is used when prior.choice=2. When prior.choice=1 an uninformative prior is used where parent combinations of all cardinalities are equally likely. The latter is equivalent to the structural prior used in the heuristic searches e.g., searchHillclimber or searchHeuristic.

Note that the network score (log marginal likelihood) of the most probable DAG is not returned as it can easily be computed using [fitAbn](#page-0-0), see examples below.

Value

An object of class abnMostprobable, which is a list containing: a matrix giving the DAG definition of the most probable posterior structure, the cache of pre-computed scores and the score used for selection.

References

Koivisto, M. V. (2004). Exact Structure Discovery in Bayesian Networks, Journal of Machine Learning Research, vol 5, 549-573.

Examples

```
## Not run:
##############################
## Example 1
##############################
## This data comes with 'abn' see ?ex1.dag.data
mydat <- ex1.dag.data[1:5000, c(1:7, 10)]
## Setup distribution list for each node:
mydists <- list(b1 = "binomial",
                p1 = "poisson",
                g1 = "gaussian",b2 = "binomial",p2 = "poisson",
                b3 = "binomial",g2 = "gaussian",g3 = "gaussian")## Parent limits, for speed purposes quite specific here:
max\_par \leftarrow list("b1" = 0,"p1" = 0,"g1" = 1,nb2" = 1,"p2" = 2,
```

```
nb3" = 3,"g2" = 3,"g3" = 2)## Now build cache (no constraints in ban nor retain)
mycache <- buildScoreCache(data.df = mydat,
                           data.dists = mydists,
                           max.parents = max_par)
## Find the globally best DAG:
mp_dag <- mostProbable(score.cache = mycache)
myres <- fitAbn(object = mp_dag,
                create.graph = TRUE)
plot(myres) # plot the best model
## Fit the known true DAG (up to variables 'b4' and 'b5'):
true_dag \leq matrix(data = 0, ncol = 8, nrow = 8)
colnames(true_dag) <- rownames(true_dag) <- names(mydists)
true_dag["p2", c("b1", "p1")] <- 1
true_dag["b3", c("b1", "g1", "b2")] <- 1
true_dag["g2", c("p1", "g1", "b2")] <- 1
true_dag["g3", c("g1", "b2")] <- 1
fitAbn(dag = true_dag,
       data.df = mydat,data.dists = mydists)$mlik
#################################################################
## Example 2 - models with random effects
#################################################################
## This data comes with abn see ?ex3.dag.data
mydat <- ex3.dag.data[, c(1:4, 14)]
mydists <- list(b1 = "binomial",
                b2 = "binomial",
                b3 = "binomial",
                b4 = "binomial")## This takes a few seconds and requires INLA:
mycache_mixed <- buildScoreCache(data.df = mydat,
                                 data.dists = mydists,
                                 group.var = "group",max.parents = 2)
## Find the most probable DAG:
mp_dag <- mostProbable(score.cache = mycache_mixed)
## and get goodness of fit:
fitAbn(object = mp_dag,
       group.var = "group")$mlik
## End(Not run)
```


Print number of observations of objects of class abnFit

Usage

```
## S3 method for class 'abnFit'
nobs(object, ...)
```
Arguments

Value

prints the number of observations of the fitted model.

Description

Probability to odds

Usage

odds(x)

Arguments

x numeric vector of probabilities with values between [0,1].

Value

numeric vector of same length as x.

Compute the odds ratio from a contingency table or a matrix.

Usage

or(x)

Arguments

x a 2x2 table or matrix.

Value

A real value.

plot.abnDag *Plots DAG from an object of class* abnDag

Description

Plots DAG from an object of class abnDag

Usage

S3 method for class 'abnDag' $plot(x, \ldots)$

Arguments

Value

Rgraphviz::plot

Examples

```
mydag <- createAbnDag(dag = ~a+b|a,
                      data.df = data.frame("a"=1, "b"=1),
                      data.dists = list(a="binomial", b="gaussian"))
plot(mydag)
```


Plot objects of class abnFit

Usage

S3 method for class 'abnFit' $plot(x, \ldots)$

Arguments

Value

a plot of the fitted model.

plot.abnHeuristic *Plot objects of class* abnHeuristic

Description

Plot objects of class abnHeuristic

Usage

```
## S3 method for class 'abnHeuristic'
plot(x, ...)
```
Arguments

Value

plot of the scores of the heuristic search.

plot.abnHillClimber *Plot objects of class* abnHillClimber

Description

Plot objects of class abnHillClimber

Usage

S3 method for class 'abnHillClimber' $plot(x, \ldots)$

Arguments

Value

plot of the consensus DAG.

plot.abnMostprobable *Plot objects of class* abnMostprobable

Description

Plot objects of class abnMostprobable

Usage

```
## S3 method for class 'abnMostprobable'
plot(x, \ldots)
```
Arguments

Value

plot of the mostprobable consensus DAG.

Print objects of class abnCache

Usage

S3 method for class 'abnCache' print(x, digits = $3, ...$)

Arguments

Value

summary statement of the class of abnCache.

Examples

```
## Subset of the build-in dataset, see ?ex0.dag.data
mydat <- ex0.dag.data[,c("b1","b2","g1","g2","b3","g3")] ## take a subset of cols
## setup distribution list for each node
mydists <- list(b1="binomial", b2="binomial", g1="gaussian",
                g2="gaussian", b3="binomial", g3="gaussian")
# Structural constraints
# ban arc from b2 to b1
# always retain arc from g2 to g1
## parent limits
max.par <- list("b1"=2, "b2"=2, "g1"=2, "g2"=2, "b3"=2, "g3"=2)
## now build the cache of pre-computed scores accordingly to the structural constraints
if(requireNamespace("INLA", quietly = TRUE)){
  # Run only if INLA is available
res.c <- buildScoreCache(data.df=mydat, data.dists=mydists,
                         dag.banned= ~b1|b2, dag.retained= ~g1|g2, max.parents=max.par)
print(res.c)
}
```


Print objects of class abnDag

Usage

S3 method for class 'abnDag' $print(x, digits = 3L, ...)$

Arguments

Value

outputs adjacency matrix and statement of the class of x.

Examples

```
mydag <- createAbnDag(dag = ~a+b|a, data.df = data.frame("a"=1, "b"=1))
print(mydag)
```
print.abnFit *Print objects of class* abnFit

Description

Print objects of class abnFit

Usage

S3 method for class 'abnFit' $print(x, digits = 3L, ...)$

Arguments

print.abnHeuristic 41

Value

prints the parameters of the fitted model.

print.abnHeuristic *Print objects of class* abnHeuristic

Description

Print objects of class abnHeuristic

Usage

S3 method for class 'abnHeuristic' $print(x, digits = 2L, ...)$

Arguments

Value

prints the best score found and the distribution of the scores.

print.abnHillClimber *Print objects of class* abnHillClimber

Description

Print objects of class abnHillClimber

Usage

```
## S3 method for class 'abnHillClimber'
print(x, digits = 3L, ...)
```
Arguments

Value

prints the consensus DAG and the class of the object.

print.abnMostprobable *Print objects of class* abnMostprobable

Description

Print objects of class abnMostprobable

Usage

S3 method for class 'abnMostprobable' $print(x, digits = 3L, ...)$

Arguments

Value

prints the mostprobable consensus DAG.

scoreContribution *Compute the score's contribution in a network of each observation.*

Description

This function computes the score's contribution of each observation to the total network score.

Usage

```
scoreContribution(object = NULL,
                         dag = NULL, data.df = NULL, data.dists = NULL,verbose = FALSE)
```
Arguments

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Details

This function computes the score contribution of each observation to the total network score. This function is available only in the mle settings. To do so one uses the $g1m$ and [predict](#page-0-0) functions. This function is an attempt to perform diagnostic for an ABN analysis.

Value

A named list that contains the scores contributions: maximum likelihood, aic, bic, mdl and diagonal values of the hat matrix.

Examples

```
## Not run:
## Use a subset of a built-in simulated data set
mydat <- ex1.dag.data[,c("b1","g1","p1")]
## setup distribution list for each node
mydists <- list(b1="binomial", g1="gaussian", p1="poisson")
## now build cache
mycache <- buildScoreCache(data.df = mydat, data.dists = mydists, max.parents = 2, method = "mle")
## Find the globally best DAG
mp.dag <- mostProbable(score.cache=mycache, score="bic", verbose = FALSE)
out <- scoreContribution(object = mp.dag)
## Observations contribution per network node
boxplot(out$bic)
## End(Not run)
```
searchHeuristic *A family of heuristic algorithms that aims at finding high scoring directed acyclic graphs*

Description

A flexible implementation of multiple greedy search algorithms to find high scoring network (DAG)

Usage

```
searchHeuristic(score.cache, score = "mlik",
                       num.searches = 1, seed = 42L, start.dag = NULL,
                       max.steps = 100,
                       algo = "hc", tabu.memory = 10, temperature = 0.9,
                       verbose = FALSE, ...)
```
Arguments

Details

This function is a flexible implementation of multiple greedy heuristic algorithms, particularly well adapted to the abn framework. It targets multi-random restarts heuristic algorithms. The user can select the num. searches and the maximum number of steps within by max. steps. The optimization algorithm within each search is relatively rudimentary.

The function searchHeuristic is different from the [searchHillClimber](#page-45-1) in the sense that this function is fully written in R, whereas the [searchHillClimber](#page-45-1) is written in C and thus expected to be faster. The function [searchHillClimber](#page-45-1) at each hill-climbing step consider every information from the pre-computed scores cache while the function [searchHeuristic](#page-42-1) performs a local stepwise optimization. This function chooses a structural move (or edge move) and compute the score's change. On this point, it is closer to the MCMCMC algorithm from Madigan and York (1995) and Giudici and Castelo (2003) with a single edge move.

If the user select random, then a random valid DAG is selected. The routine used favourise low density structure. The function implements three algorithm selected with the parameter algo: hc, tabu or sa.

If algo=hc: The Hill-climber algorithm (hc) is a single move algorithm. At each Hill-climbing step within a search an arc is attempted to be added. The new score is computed and compared to the previous network's score.

If algo=tabu: The same algorithm is as with hc is used, but a list of banned moves is computed. The parameter tabu.memory controls the length of the tabu list. The idea is that the classical Hillclimber algorithm is inefficient when it should cross low probability regions to unblock from a local maximum and reaching a higher score peak. By forcing the algorithm to choose some not already used moves, this will force the algorithm to escape the local maximum.

If algo=sa: This variant of the heuristic search algorithm is based on simulated annealing described by Metropolis et al. (1953). Some accepted moves could result in a decrease of the network score. The acceptance rate can be monitored with the parameter temperature.

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Value

An object of class abnHeuristic (which extends the class abnLearnd) and contains list with entires:

dags a list of DAGs

scores a vector giving the network score for the locally optimal network for each search

detailed.score a vector giving the evolution of the network score for the all the random restarts

score a number giving the network score for the locally optimal network

score.cache the pre-computed cache of scores

num.searches a numeric giving the number of random restart

max.steps a numeric giving the maximal number of optimization steps within each search

algorithm a character for indicating the algorithm used

References

Heckerman, D., Geiger, D. and Chickering, D. M. (1995). Learning Bayesian networks: The combination of knowledge and statistical data. *Machine Learning*, 20, 197-243. Madigan, D. and York, J. (1995) "Bayesian graphical models for discrete data". International Statistical Review, 63:215232. Giudici, P. and Castelo, R. (2003). "Improving Markov chain Monte Carlo model search for data mining". Machine Learning, 50:127158. Metropolis, N., Rosenbluth, A. W., Rosenbluth, M. N., Teller, A. H., & Teller, E. (1953). "Equation of state calculations by fast computing machines". The journal of chemical physics, 21(6), 1087-1092.

Examples

```
## Not run:
##############################################
## example: use built-in simulated data set
##############################################
mydat <- ex1.dag.data ## this data comes with abn see ?ex1.dag.data
## setup distribution list for each node
mydists<-list(b1="binomial", p1="poisson", g1="gaussian", b2="binomial",
              p2="poisson", b3="binomial", g2="gaussian", b4="binomial",
              b5="binomial", g3="gaussian")
mycache <- buildScoreCache(data.df = mydat, data.dists = mydists, max.parents = 2)
## Now peform 10 greedy searches
heur.res <- searchHeuristic(score.cache = mycache, data.dists = mydists,
                            start.dag = "random", num.searches = 10,
                            max.steps = 50## Plot (one) dag
plotAbn(heur.res$dags[[1]], data.dists = mydists)
## End(Not run)
```


Find high scoring network (DAG) structures using a random re-starts greedy hill-climber heuristic search.

Usage

```
searchHillClimber(score.cache, score = "mlik", num.searches = 1, seed = 42,
                    start.dag = NULL, support.threshold = 0.5, timing.on = TRUE,
                         dag.retained = NULL, verbose = FALSE, ...)
```
Arguments

Details

The procedure runs a greedy hill-climbing search similar, but not identical, to the method presented initially in Heckerman et al. 1995. (Machine Learning, 20, 197-243). Each search begins with a randomly chosen DAG structure where this is constructed in such a way as to attempt to choose a DAG uniformly from the vast landscape of possible structures. The algorithm used is as follows: given a node cache (from [buildScoreCache](#page-0-0), then within this set of all allowed local parent combinations, a random combination is chosen for each node. This is then combined into a full DAG, which is then checked for cycles, where this check iterates over the nodes in a random order. If all nodes have at least one child (i.e., at least one cycle is present), then the first node examined has all its children removed, and the check for cycles is then repeated. If this has removed the only cycle present, then this DAG is used at the starting point for the search, if not, a second node is chosen (randomly) and the process is then repeated until a DAG is obtained.

The actual hill-climbing algorithm itself differs slightly from that presented in Heckerman et al. as a full cache of all possible local combinations are available. At each hill-climbing step, everything in the node cache is considered. In other words, all possible single swaps between members of cache currently present in the DAG and those in the full cache. The single swap, which provides the greatest increase in goodness of fit is chosen. A single swap here is the removal or addition of any one node-parent combination present in the cache while avoiding a cycle. This means that as well as all single arc changes (addition or removal), multiple arc changes are also considered at each same step, note however that arc reversal in this scheme takes two steps (as this requires first removal of a parent arc from one node and then addition of a parent arc to a different node). The original algorithm perturbed the current DAG by only a single arc at each step but also included arc reversal. The current implementation may not be any more efficient than the original but is arguably more natural given a pre-computed cache of local scores.

A start DAG may be provided in which case num.searches must equal 1 - this option is really just to provide a local search around a previously identified optimal DAG.

This function is designed for two different purposes: i) interactive visualization; and ii) longer batch processing. The former provides easy visual "eyeballing" of data in terms of its majority consensus network (or similar threshold), which is a graphical structure which comprises of all arcs which feature in a given proportion (support.threshold) of locally optimal DAGs already identified during the run. The general hope is that this structure will stabilize - become fixed - relatively quickly, at least for problems with smaller numbers of nodes.

Value

A list with entries:

init.score a vector giving network score for initial network from which the search commenced

final.score a vector giving the network score for the locally optimal network

init.dag list of matrices, initial DAGs

final.dag list of matrices, locally optimal DAGs

consensus a matrix holding a binary graph, not necessary a DAG!

support.threshold percentage supported used to create consensus matrix

References

Lewis, F. I., and McCormick, B. J. J. (2012). Revealing the complexity of health determinants in resource poor settings. *American Journal Of Epidemiology*. DOI:10.1093/aje/KWS183).

Simulate data from a fitted additive Bayesian network.

Usage

```
simulateAbn(
 object = NULL,run.simulation = TRUE,
 bugsfile = NULL,
 n.chains = 10L,
 n.addapt = 1000L,n.thin = 100L,
 n.iter = 10000L,
  seed = 42L,
 verbose = FALSE,
  debug = FALSE
)
```
Arguments

Value

data.frame

See Also

[makebugs](#page-0-0)

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Examples

```
df <- FCV[, c(12:15)]mydists <- list(Outdoor="binomial",
                Sex="multinomial",
                GroupSize="poisson",
                Age="gaussian")
## buildScoreCache -> mostProbable() -> fitAbn()
suppressWarnings({
  mycache.mle <- buildScoreCache(data.df = df, data.dists = mydists, method = "mle",
                                 adj.vars = NULL, cor.vars = NULL,
                                 dag.banned = NULL, dag.retained = NULL,
                                 max.parents = 1,
                                 which.nodes = NULL, defn.res = NULL)
}) # ignore non-convergence warnings
mp.dag.mle <- mostProbable(score.cache = mycache.mle, verbose = FALSE)
myres.mle <- fitAbn(object = mp.dag.mle, method = "mle")
myres.sim <- simulateAbn(object = myres.mle,
                             run.simulation = TRUE,
                             bugsfile = NULL,
                             verbose = FALSE)
str(myres.sim)
prop.table(table(myres.sim$Outdoor))
prop.table(table(df$Outdoor))
```


Description

Provided with node names, returns an abnDAG. Arc density refers to the chance of a node being connected to the node before it.

Usage

```
simulateDag(node.name, data.dists = NULL, edge.density = 0.5, verbose = FALSE)
```
Arguments

Details

This function generates DAGs by sampling triangular matrices and reorder columns and rows randomly. The network density (edge.density) is used column-wise as binomial sampling probability. Then the matrix is named using the user-provided names.

Value

object of class abnDag consisting of a named matrix, a named list giving the distribution for each node and an empty element for the data.

Examples

```
simdag <- simulateDag(node.name = c("a", "b", "c", "d"),
                     edge.density = 0.5,
                     data.dists = list(a = "gaussian",b = "binomial",
                                       c = "poisson",
                                       d = "multinomial"))
## Example using Ozon entries:
dist <- list(Ozone="gaussian", Solar.R="gaussian", Wind="gaussian",
            Temp="gaussian", Month="gaussian", Day="gaussian")
out <- simulateDag(node.name = names(dist), data.dists = dist, edge.density = 0.8)
plot(out)
```
skewness *Computes skewness of a distribution*

Description

Computes skewness of a distribution

Usage

```
skewness(x)
```
Arguments

x a numeric vector

Value

integer

Prints summary statistics from an object of class abnDag

Usage

```
## S3 method for class 'abnDag'
summary(object, ...)
```
Arguments

Value

List with summary statistics of the DAG.

Examples

```
mydag <- createAbnDag(dag = ~a+b|a, data.df = data.frame("a"=1, "b"=1))
summary(mydag)
```
summary.abnFit *Print summary of objects of class* abnFit

Description

Print summary of objects of class abnFit

Usage

S3 method for class 'abnFit' summary(object, digits = 3L, ...)

Arguments

Value

prints summary statistics of the fitted model.

```
summary.abnMostprobable
```
Print summary of objects of class abnMostprobable

Description

Print summary of objects of class abnMostprobable

Usage

```
## S3 method for class 'abnMostprobable'
summary(object, ...)
```
Arguments

Value

prints the mostprobable consensus DAG and the number of observations used to calculate it.

toGraphviz *Convert a DAG into graphviz format*

Description

Given a matrix defining a DAG create a text file suitable for plotting with graphviz.

Usage

```
toGraphviz(dag,
```

```
data.df=NULL,
data.dists=NULL,
group.var=NULL,
outfile=NULL,
directed=TRUE,
verbose=FALSE)
```
toGraphviz 53

Arguments

Details

Graphviz (<https://www.graphviz.org>) is a visualisation software developed by AT&T and freely available. This function creates a text representation of the DAG, or the undirected graph, so this can be plotted using graphviz. The R package, Rgraphviz (available through the Bioconductor project <https://www.bioconductor.org/>) interfaces R and the working installation of graphviz.

Binary nodes will appear as squares, Gaussian as ovals and Poisson nodes as diamonds in the resulting graphviz network diagram. There are many other shapes possible for nodes and numerous other visual enhancements - see online graphviz documentation.

Bespoke refinements can be added by editing the raw outfile produced. For full manual editing, particularly of the layout, or adding annotations, one easy solution is to convert a postscript format graph (produced in graphviz using the -Tps switch) into a vector format using a tool such as pstoedit (<http://www.pstoedit.net/>), and then edit using a vector drawing tool like xfig. This can then be resaved as postscript or pdf thus retaining full vector quality.

Value

Nothing is returned, but a file outfile written.

Author(s)

Fraser Iain Lewis

Marta Pittavino

Examples

```
## On a typical linux system the following code constructs a nice
## looking pdf file 'graph.pdf'.
## Not run:
## Subset of a build-in dataset
mydat <- ex0.dag.data[,c("b1","b2","b3","g1","b4","p2","p4")]
```
setup distribution list for each node

```
mydists <- list(b1="binomial", b2="binomial", b3="binomial",
               g1="gaussian", b4="binomial", p2="poisson",
                               p4="poisson")
## specify DAG model
mydag <- matrix(c( 0,1,0,0,1,0,0, #
                    0,0,0,0,0,0,0, #
                    0,1,0,0,1,0,0, #
                    1,0,0,0,0,0,1, #
                    0,0,0,0,0,0,0, #
                    0,0,0,1,0,0,0, #
                    0,0,0,0,1,0,0 #
), byrow=TRUE, ncol=7)
colnames(mydag) <- rownames(mydag) <- names(mydat)
## create file for processing with graphviz
outfile <- paste(tempdir(), "graph.dot", sep="/")
toGraphviz(dag=mydag, data.df=mydat, data.dists=mydists, outfile=outfile)
## and then process using graphviz tools e.g. on linux
if(Sys.info()[["sysname"]] == "Linux" & interactive())system(paste( "dot -Tpdf -o graph.pdf", outfile))
 system("evince graph.pdf")
}
## Example using data with a group variable where b1<-b2
mydag \leq matrix(c(0,1, 0,0), byrow=TRUE, ncol=2)
colnames(mydag) <- rownames(mydag) <- names(ex3.dag.data[,c(1,2)])
## specific distributions
mydists <- list(b1="binomial", b2="binomial")
## create file for processing with graphviz
outfile <- paste0(tempdir(), "/graph.dot")
toGraphviz(dag=mydag, data.df=ex3.dag.data[,c(1,2,14)], data.dists=mydists,
          group.var="group",
          outfile=outfile, directed=FALSE)
## and then process using graphviz tools e.g. on linux:
if(Sys.info()[["sysname"] = "Linux" & interactive())pdffile <- paste0(tempdir(), "/graph.pdf")
 system(paste("dot -Tpdf -o ", pdffile, outfile))
 system(paste("evince ", pdffile, " &")) ## or some other viewer
}
```
End(Not run)

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